Voltammetry: an in situ multi-analyte tool to monitor the health of estuaries

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Outline

In situ techniques are necessary to study the environment – voltammetry as a good *non-selective* analytical method Show capability in a variety of estuarine environments **Inland Bays – lab equipment brought to the field Chesapeake Bay – field equipment Field equipment on moorings:** Salt marsh / sediment work **Delaware Bay**

A story on O₂, Fe, S biogeochemistry coupled with physical forcing

PEEK & Glass encased electrodes in marine epoxy

100 μm diameter Au wire



Water column / vent working electrode

Sediment working electrode

Plate with Hg film

O₂, Fe²⁺, Mn²⁺, H₂S, H₂O₂, I⁻, S_x²⁻, S₂O₃²⁻, FeS_{aq}, Fe(III) are all measurable in one scan, if present

Tested to 2700 m and 120 °C Solid state (micro)electrodes for the analysis of biologically relevant compounds and ions Chemistry Drives Biology Rationale for design and use

Fine scale resolution - mm in sediments; micrometer in biofilms and mats

determine sediment heterogeneity vs. homogeneity use to prospect for life forms and understand ecosystem health Use in sedimentary porewaters of bays, oceans and lakes in water column; e.g., Chesapeake Bay, Black Sea at Hydrothermal Vents, Yellowstone hot springs, in corrosion studies

VOLTAMMETRY I vs **E** plots [similar to A vs λ plots]

Vertical lines indicate the half-wave potential for the reduction of each analyte at the Au/Hg electrode

Potential scans and Hg tip prevent (bio)fouling



Detection limits 3 μM for O₂, 5 μM Mn²⁺ 10 μM Fe²⁺ 0.1 μM H₂S

Multi-analyte sensor

No standards for Fe(III) and FeS

Luther et al, 2008

Where is the oxygen?

Oxic

concentration

 O_2 - oxygen

 $N \emptyset_3^-$ - nitrate

Mn²⁺ -)manganese

Fe²⁺ - iron

 $H_2S - hydrogen sulfide$



Due to Organic matter decomposition by oxidants [oxygen, nitrate, manganese and iron oxides (rust), sulfate]

Suboxic – no oxygen and no hydrogen sulfide

 $Fe^{2+} + H_2S \rightarrow FeS + 2 H^+$

 $FeS + H_2S \rightarrow FeS_2 + H_2$

fools' gold

Anoxic – no oxygen BUT also rich in ammonia and phosphate (nutrients)



In situ comparison of O₂ Clark vs voltammetric Au/Hg in sediments from a ROV



Real time voltammetry of porewaters





Free Benthic Lander for Depth Profiles and Flux Measurements



Benthic chamber with Au/Hg microelectrode



SOUTH

GEORGIA

-Remote micromanipulator Potentiostat: ISEA IITM AIS Analytical Instrument Systems, Inc.

Cable for all 4 Au/Hg electrodes

Tracer injection device



SEDIMENT DIAGENESIS



pH reflects processes

> O₂, NO₃⁻ electron acceptor; lowers pH

Mn²⁺ inc.; MnO₂ electron acceptor; raises pH

Ca(Mn)CO₃ precipitation; lowers pH

 $\begin{array}{rl} 236 \ MnO_2 \ + \ C_{106}H_{263}O_{110}N_{16}P \ + \ 364 \ H^+ \\ \rightarrow \ 8 \ N_2 \ + \ 106 \ HCO_3^- \ + \ HPO_4^{\ 2-} \ + \ 236 \\ Mn^{2+} \ + \ 636 \ H_2O \end{array}$

Luther et al, 1999

Tourquay Canal DE – July 10, 2000 benthic processes at their worst?





Close-up



Electrodes on wire for *in situ* work

End of canal

Fish found at end of canal at control sites, brought in with the tide



Blue crabs stressed in early August 5, 2004 –

there were small amounts of oxygen in the surface waters along with H₂S









Delaware's Inland Bays

Bald Eagle Creek area residence time 90 days

Only One Inlet

System with significant eutrophication

Torquay Canal / Bald Eagle Creek Sites



Nutrients from

Waste water treatment plant

Golf course

Torquay Canal / Bald Eagle Creek Sites



sites 2 and 9 are larger than a FOOTBALL FIELD IN AREA

Torquay canal control sites = 1, 5

Circles indicate holes in this ecosystem with depths of 5 meters. Squares have normal depth of 2 m

In situ Voltammograms



Temperature change from Spring to Summer, 2001 SITE #2



T-5.01, low tide,

- ---- T-5.11, high tide, cold and windy last week
- ---- T-5.24, two day's rain and thunderstorm
- T-6.13, high tide, several days warm
- T-6.27, low tide, thunder storm and rain 4 days ago
- T-7.12, high tide, strong wind overturns water

2 m mixed layer depth in summer STRONG PHYSICAL **STRATIFICATION !**

STRONG WINDS CAN OVERTURN THIS !

Mixed layer changes with season in 2001 for hole at site 2

O₂ change over time at Site #2, Tourquay Canal

H₂S change over time at Site #2, Tourquay Canal



Strong PHYSICAL AND CHEMICAL stratification. Mixed layer ~ 2 meters

Sulfide change with season in bottom waters at 4.5 meter depth.



Sulfide levels are among the highest reported in anoxic basins

Luther et al, 2004; Ma et al 2006a, b

Tourquay Canal after event #2 – bottom water overturn



 $\begin{array}{l} \mbox{Fe}^{2+} \mbox{ is ND-10 } \mu M \mbox{ (below Au/Hg detection limit); FeS}_{aq} \mbox{ is present;} \\ \mbox{ } Mn^{2+} \mbox{ is } < 0.2 \ \mu M \end{array}$

Fe catalytic cycle

 $HS^- → HS + e^- \qquad E^0 = -1.08 V$ $O_2 + e^- → O_2^- \qquad E^0 = -0.16 V$ $O_2 + HS^- → HS + O_2^- \qquad E^0 = -1.24 V \text{ (overall)}$ Reaction is thermodynamically unfavorable!Thus, Oxygen does NOT directly oxidize hydrogen sulfide!

BUT

 $2 \operatorname{Fe}^{2+} + \operatorname{O}_2 + 4 \operatorname{H}^+ \rightarrow 2 \operatorname{Fe}^{3+} + 2 \operatorname{H}_2 \operatorname{O}_2$ $2 \operatorname{Fe}^{3+} + \operatorname{HS}^- \rightarrow 2 \operatorname{Fe}^{2+} + \operatorname{S}^0 + \operatorname{H}^+$

S° concentrations were as high as 30 micromolar and should be added to the H₂S data for total reduced sulfur.

Thermodynamics and kinetics of Fe²⁺ and Mn²⁺ oxidation in freshwater (25 °C) at O₂ saturation predicts abiotic reaction





Ma et al, 2006a, b



Submersible Electrochemical Analyzer on Deck

SeaBird MicroCat CTD

Voltammetric electrode holder for 4 working electrodes

pH probe

Extra temperature sensor



Deploying Submersible Electrochemical Analyzer







Stand Alone

July 28, 2002 @ 0900 – high slack water



Upcast vs Downcast July 28, 2002 @ 1230 – mid ebb tide



July 28 2002 @ 1600 low slack water





Slack waters allow for the suboxic zone to setup again

Moving waters destroy the suboxic zone and cause more oxidation

Lewis et al, 2007

Skidaway Island Salt Marsh Environmental Research Facility (SERF)







Difference between high and low tide = 2.5 m



Remote Sensing of Salt Marsh Geochemistry











In Situ Measurements in Mud Flat Sediments



Average Fe²⁺ and $\Sigma H_2 S$ Concentrations Measured *In Situ* as a Function of Depth



TC = Tidal Cycle

Taillefert et al. (2007)

Conclusions

Ex situ core measurements cannot capture the dynamics of biogeochemical processes in sediments affected by tidal forcing

Overall, mud flat sediments are dominated by sulfate reduction, while creek bank sediments are dominated by metal reduction Hydrostatic pressure is higher in creek bank than in mud flat sediments Creek bank sediments are more oxidized (brownish sediments)

Voltammetry can be used *in situ* to determine the effect of tidal forcing on biogeochemical processes

Moorings with physical, chemical and biological sensors

Measures salinity, temperature, O₂, fluorescence, transmissometry, light irr<u>adiance – (data to shore via SWAP</u>)



Data Transmission via

SWAP: Shipboard Wireless Access Protocol



Ship to ship Ship to shore Shore to shore with 2.4 gHz radio About 15 km

Delaware Bay Mooring Location



Time course data for 1 month; 0.5 hr data collection



Higher O₂ / higher productivity

Lower O₂ so lower productivity / higher respiration or photoinhibition?

Productivity and Respiration model from Chapra (1997)



Productivity and Respiration model from Chapra (1997) P (productivity) = $r_{oa}G_{max}1.066^{T-20} \Phi_{l}a$ R (respiration) = $r_{oa} k_{ra}1.08^{T-20}a$

Physical processes accounted for with wind speed

 $r_{oa} = O_2$ generated from unit mass of plant biomass produced (mg-O₂ / mg-Chla) $G_{max} =$ maximum plant growth rate for optimal light and excess nutrient conditions

T = water temperature (° **C**)

a = concentration of plant biomass (mg-Chla m⁻³)

 Φ_{l} = attenuation of growth due to light

k_{ra} = **plant respiration rate**

The model is not fully constrained but can be

 r_{oa} and G_{max} are unknown conversion factors from chl a to O_2 and light to growth

These can be measured or estimated with an *in situ* device known as the Fluorescence Induction Relaxation system (FIRe) - (Gorbunov and Falkowski, 2004)

FIRe measures the variable fluorescence of the photosystem center II, and thus is an indicator of how efficient a photosynthetic organism can utilize light energy to fix inorganic carbon and produce O_2 .

Oxygenic photosynthesis



IMPACT OF ELECTRODES ON THE FIELD

Multi-analyte Environmental chemistry tool

Sediment Diagenesis better described with finer resolution

Describe ecosystem health - why do fish kills and HABs occur? Benthic – pelagic coupling;

Must combine several tools/data to understand physics, chemistry and their role on biology: need newer biological sensors (FIRe) to Assess planktonic health at molecular level

Increased collaboration with government officials and industry with recommendations for improvement

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University of Delaware Marine Campus in Lewes



Voltammetric O₂ data



Monitoring Wells to Measure Water Levels in Sediments



Taillefert et al. (2007)



Tidal Advection in Sediments

- Tidal forcing affects porewater advection over 30 cm only
- Compared to mud flat sediments, creek bank sediments display:
 - higher hydrostatic pressure
 - faster advection rates
- Water infiltrates mud flat sediments from the surface, while deep wells fill first in creek bank sediments

Taillefert et al. (2007) Geochem. Trans.



Time since started - [hh:mm]

In Situ Measurements in Creek Bank Sediments



 At ebb tide, species diffuse away or are oxidized by O_{2(aq)} that can penetrate from the overlying waters.



Seasonal Variations in Biogeochemical Processes at SAT2 and SAT4

SAT 2 (freshwater

SAT 4 (marine side)



Fe(III) production and reduction to Fe²⁺ are more prevalent in SAT1 and SAT 2 sediments

SO₄²⁻ reduction becomes more prevalent in SAT 4, SAT5, and SAT 6 sediments during late summer and fall only: based on porewater data